Synthesis, Structure, and Cytotoxicity of *O*-[3-(5-Tetrazolyl)propyl]oximes

K. Rubina, E. Abele, S. Belyakov, I. Shestakova, and Yu. Popelis

Latvian Institute of Organic Chemistry, Riga, LV-1006 Latvia e-mail: kira@osi.lv

Received February 14, 2005

Abstract—A two-stage method was developed for the conversion of phenyl- and pyridylaldoximes and ketoximes into the corresponding O-[3-(5-tetrazolyl)propyl]oximes. The structure of the sodium salt of 2-acetylpyridine O-[3-(5-tetrazolyl)propyl]oxime was established X-ray diffraction analysis. This compound was shown to possess a high cytotoxic activity in vitro.

DOI: 10.1134/S1070428006050149

Tetrazole derivatives containing oxygen and nitrogen [1–4] are of interest as biologically active compounds [5]. Recently their action was extensively investigated with respect to the cardiovascular system [6–11]. Besides the tetrazole derivatives are used as antihistaminic [12], antiphlogistic [13], and antiulcer [14] preparations. Thiobis(alkylene-5-tetrazoles)[15] and 1-(2-ethylthio-6-aminomethylbenzothiazolyl)-2-phenyl-1,3,4,5-tetrazole [16] were found to possess antitumor activity. However the oxime tetrazole derivatives are poorly understood. It was only established that carbamates of tetrazolyl-(alkylene)oximes exhibited the pesticide activity [17]. As far as we know the cytotoxicity of tetrazole oxime derivatives has not been formerly investigated.

The goal of this study was the synthesis, structure identification, and the study of the cytotoxic activity of new *O*-[3-(5-tetrazolyl)propyl]oximes.

A method for preparation of these oximes **IX–XII** from aldoximes and ketoximes **I–IV** was developed (see the scheme).

Initial oximes I–IV in the phase-transfer catalytic systems NCCH₂CH₂CH₂Br–solid KOH–18-crown-6–toluene or NCCH₂CH₂CH₂Br–10% aqueous NaOH–Oct₄NBr–toluene afforded *O*-(3-cyanopropyl)oximes V–VIII in 30–97% yields. The subsequent reaction of oximes V–VIII with azidotrimethylsilane in the presence of Bu₂SnO [18] resulted in the formation of tetrazoles IX–XII in 39–65% yields.

2-Acetylpyridine *O*-[3-(5-tetrazolyl)propyl]oxime (**XII**) was isolated as a sodium salt crystal hydrate. According to X-ray diffraction data (Fig. 1) three water molecules are involved in the coordination bonds with the sodium ion. The values of the principal bond lengths and bond angles in the anion of salt **XII** are given in Table 1; there are also listed the lengths of the coordination bonds formed by Na⁺ ion.

In the crystal structure of compound **XII** an intricate system exists of intermolecular hydrogen bonds. As seen from Table 2, these bonds are somewhat longer than the statistical average values which for intermolecular

Scheme.

Ar = Ph(I, II, V, VI, IX, X), 2-pyridyl(III, IV, VII, VIII, XI, XII); R = H(I, III, V, VII, IX, XI), Me(II, IV, VI, VIII, X, XII).

736 RUBINA et al.

Table 1. Interatomic distances (l) and bond angles (ω) in the structure of 2-acetylpyridine O-[3-(5-tetrazolyl)propyl]oxime sodium salt crystal hydrate (**XII**)

sociali sait orystai nyarate (ATT)						
Bond	l, A	Angle	ω, deg			
N^3-N^4	1.349(3)	$N^4N^3N^2$	109.1(2)			
N^3-N^2	1.307(3)	$N^{I\theta}O^9C^8$	109.2(2)			
$O^9 - N^{10}$	1.399(3)	$C^6C^5N^4$	125.2(2)			
O^9 – C^8	1.438(3)	$C^6C^5N^I$	123.7(2)			
C^5-C^6	1.490(3)	$N^4C^5N^I$	111.1(2)			
C^5-N^4	1.332(3)	$\mathbf{C}^{5}\mathbf{C}^{6}\mathbf{C}^{7}$	114.2(2)			
C^5-N^I	1.331(3)	$O^9N^{I0}C^{II}$	111.6(2)			
$\mathbf{C}^6 - \mathbf{C}^7$	1.514(3)	$N^3N^4C^5$	105.2(2)			
N^{I0} – C^{II}	1.276(3)	$C^5N^IN^2$	105.2(2)			
$N^{I}-N^{2}$	1.344(3)	$C^{12}N^{13}C^{14}$	117.8(3)			
N^{13} – C^{12}	1.352(4)	$N^3N^2N^I$	109.5(2)			
N^{13} – C^{14}	1.335(4)	$\mathbf{C}^{6}\mathbf{C}^{7}\mathbf{C}^{8}$	112.5(2)			
$\mathbf{C}^7 - \mathbf{C}^8$	1.514(3)	$O^9C^8C^7$	106.8(2)			
\mathbf{C}^{II} – \mathbf{C}^{I2}	1.487(4)	$N^{I0}C^{II}C^{I2}$	116.1(2)			
\mathbf{C}^{II} – \mathbf{C}^{I8}	1.499(4)	$N^{I\theta}C^{II}C^{I8}$	123.1(3)			
$C^{12}-C^{17}$	1.386(4)	$\mathbf{C}^{12}\mathbf{C}^{11}\mathbf{C}^{18}$	120.8(3)			
C^{14} – C^{15}	1.397(5)	$N^{I3}C^{I2}C^{II}$	117.0(2)			
$C^{17} - C^{16}$	1.368(6)	$N^{I3}C^{I2}C^{I7}$	121.5(3)			
$C^{16} - C^{15}$	1.355(7)	$\mathbf{C}^{II}\mathbf{C}^{I2}\mathbf{C}^{I7}$	121.5(3)			
$Na^{I}\cdots O^{I}$	2.405(2)	$N^{13}C^{14}C^{15}$	122.6(4)			
$Na^{I} \cdots O^{2}$	2.318(2)	$\mathbf{C}^{12}\mathbf{C}^{17}\mathbf{C}^{16}$	120.1(4)			
$Na^{I} \cdots O^{3}$	2.298(2)	$C^{17}C^{16}C^{15}$	118.9(3)			
$Na^{I}\cdots N^{I0}$	2.506(3)	$\mathbf{C}^{14}\mathbf{C}^{15}\mathbf{C}^{16}$	119.2(4)			
$Na^{I}\cdots N^{I3}$	2.471(3)					

hydrogen bonds of OH···N and OH···O types equal 2.79 and 2.72 A respectively [19]. The projection of the crystal structure is given on Fig. 2.

We investigated some of compounds obtained for their cytotoxic activity in vitro. The toxicity evaluation of

Table 2. Parameters of hydrogen bonds in the crystal structure of 2-acetylpyridine *O*-[3-(5-tetrazolyl)propyl]oxime sodium salt crystal hydrate (**XII**)

Bond D–H···A	Length of H-bond D···A, Å	Distance D···A, A	Angle, DH···Å, deg	Position of atom A
O^{I} $-H^{IA}$ $\cdots N^{3}$	2.893(3)	2.08(3)	175(3)	1-x, -y, 1-z
O^{I} $-H^{IB}N^{4}$	2.804(3)	1.91(4)	174(3)	-1 + x, y, z
O^2 - H^{2A} ··· O^I	2.836(3)	2.04(4)	167(3)	-x, $-1-y$, $1-z$
$O^2-H^{2B}\cdots N^I$	2.821(3)	2.00(5)	167(4)	x, -1+y, z
$O^3-H^{3A}\cdots N^2$	2.792(3)	1.89(4)	174(3)	-1+x, -1+y, z
O^3 – H^{3B} ··· O^2	2.883(3)	2.15(5)	153(4)	-x, -1-y, 1-z

compounds VI, IX, X, and XII on a cell line NIH 3T3 (mice embryos fibroblasts) showed that acetophenone O-(3-cyanopropyl)oxime (VI) is nontoxic (LD₅₀ > 2000 mg/kg), and the toxicity grew in the tetrazoles serried $XII \le X \le IX$ (LD₅₀ in the range 191–717 mg/kg). The concentration of compounds ensuring the 50% cell loss in vitro (IC₅₀) was evaluated on five cell lines: HT-1080 (human fibrosarcoma), MG-22A (hepatoma of mice), B16 (melanoma of mice), and Neuro 2A (neuroblastoma of mice), and also on normal cell lines BHK-21 (kidney cells of golden hamster) and NIH 3T3 (mice embryos fibroblasts). The experiments demonstrated that acetophenone cyanooxime VI did not exhibit the cytotoxic activity. The raising of the compound hydrophilism by introducing a tetrazole ring in acetophenone O-[3-(5tetrazolyl)propyl]oxime (X) led to the appearance of cytotoxicity: IC₅₀ was from 3.5 (HT-1080) to 28 µg/ml (Neuro 2A). Consequently, sodium salt XII soluble in water possesses high cytotoxic activity for all cell lines, both cancerous and normal [for instance, IC₅₀ 1.4 (HT-1080) and 0.9 μg/ml (BHK-21)]. Certain selectivity of cytotoxic action was observed for benzaldehyde O-[3-

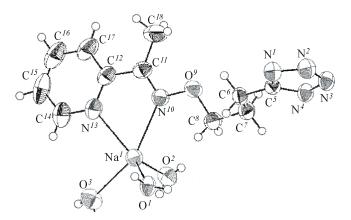


Fig. 1. Spatial model of the complex of 2-acetylpyridine *O*-[3-(5-tetrazolyl)propyl]oxime (**XII**).

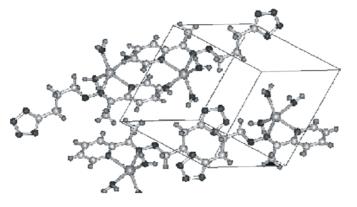


Fig. 2. Projection of the crystalline structure of the complex of 2-acetylpyridine *O*-[3-(5-tetrazolyl)propyl]oxime (**XII**).

(5-tetrazolyl)propyl]oxime (IX). Its cytotoxicity for the line HT-1080 was 3.5 μ g/ml, for NIH 3T3, 114 μ g/ml.

EXPERIMENTAL

¹H and ¹³C NMR spectra were registered on a spectrometer Varian 200 Mercury at operating frequencies 200.06 and 50.31 MHz in CDCl₃, internal reference HMDS. Mass spectra were registered on a GC-MS HP 6890 instrument with ionizing electrons energy 70 eV. GLC was performed on a chromatograph Chrom-5 equipped with a flame ionization detector, a glass column (1200×3 mm), stationary phase 5% OV-101 on Chromosorb W-HP (80–100 mesh), carrier gas nitrogen, flow rate 60 cm³/min. Oven temperature was varied in the range 180-250°C depending on the reaction mixture composition. Commercial dibutyltin oxide, 1-bromo-3cyanopropane, azidotrimethylsilane, and 18-crown-6 (Acros) were used without additional purification. Oximes I-IV were obtained from the corresponding aldehydes and ketones [20].

Benzaldehyde O-(3-cyanopropyl)oxime (V). To a suspension of 1.35 g (10 mmol) of benzaldehyde oxime, 1.68 g (30 mmol) of solid, finely dispersed KOH, and 264 mg (1 mmol) of 18-crown-6 in 10 ml of toluene was added at vigorous stirring 1.48 g (10 mmol) of 1-bromo-3-cyanopropane. The reaction continued for 5 h at room temperature (GLC monitoring). Then the reaction mixture was filtered and evaporated. The reaction product was isolated by column chromatography on silica gel, eluent toluene (R_f 0.4). Yield 1.8 g (96%). ¹H NMR spectrum, δ, ppm: 2.10 m (2H, CH₂), 2.48 t (2H, OCH₂, J 7.2 Hz), 4.25 t (2H, CH₂CN, J 6.0 Hz), 7.36 m (3H, H^{3-5}), 7.56 m (2H, $H^{2,6}$), 8.07 s (1H, CH). ¹³C NMR spectrum, δ , ppm: 14.1 (CH₂), 25.4 (OCH₂), 71.4 (<u>C</u>H₂C), 119.3, 127.0, 128.7, 131.9, 149.2. Mass spectrum, m/z $(I_{\text{rel}}, \%)$: 188 (22) M^+ , 187 (28), 157 (26), 131 (15), 120 (25), 105 (41), 104 (88), 94 (18), 89 (21), 78 (26), 77 (100), 65 (27), 51 (58). Found, %: C 70.39; H 6.43; N 14.78. C₁₁H₁₂N₂O. Calculated, %: C 70.19; H 6.43; N 14.88.

Acetophenone *O*-(3-cyanopropyl)oxime (VI) was similarly obtained within 4 h. The reaction mixture was filtered and used in the next stage of the synthesis without isolation of the product. Mass spectrum, m/z (I_{rel} , %): 202 (31) M^+ , 20 (28), 171 (58), 134 (32), 117 (30), 118 (43), 106 (17), 104 (73), 94 (10), 78 (26), 77 (100), 51 (25).

2-Pyridinealdehyde O-(3-cyanopropyl)oxime (VII). To a suspension of 0.84 g (6.7 mmol)of

2-pyridinealdehyde oxime (**III**), 0.183 g (0.34 mmol) of tetraoctylammonium bromide, 5.4 ml of 10% aqueous NaOH in 7 ml of toluene was added 1.02 g (6.7 mmol) of 1-bromo-3-cyanopropane, the mixture was stirred for 5 h at 100°C, then filtered, the filtrate was evaporated on the rotary evaporator. The residue was purified by column chromatography (eluent ethyl acetate, R_f 0.45). Yield 0.38 g (30%). ¹H NMR spectrum, δ, ppm: 2.10 m (2H, CH₂), 2.50 t (2H, OCH₂, *J* 7.6 Hz), 4.32 t (2H, CH₂CN, *J* 5.8 Hz), 7.27 m (1H, H⁵), 7.73 m (2H, H^{3,4}), 8.17 s (1H, CH), 8.61 m (1H, H⁶). ¹³C NMR spectrum, δ, ppm: 14.0 (CH₂), 25.3 (OCH₂), 71.9 (<u>C</u>H₂C), 119.2, 121.1, 124.1, 136.5, 149.7, 149.9, 151.3. Mass spectrum, m/z (I_{rel} , %): 189 (4) M^+ , 122 (12), 106 (19), 92 (17), 79 (100), 78 (62), 65 (10), 51 (30).

2-Acetylpyridine *O*-(**3-cyanopropyl)oxime** (VIII) was prepared as oxime V in 3 h. The residue was purified by column chromatography (eluent ethyl acetate, R_f 0.75). Yield 1.96 g (97%). ¹H NMR spectrum, δ , ppm: 2.10 m (2H, CH₂), 2.33 s (3H, Me), 2.50 t (2H, OCH₂, J 7.2 Hz), 4.33 t (2H, CH₂CN, J 5.6 Hz), 7.23 m (1H, H⁵), 7.66 m (1H, H⁴), 7.88 m (1H, H³), 8.59 m (1H, H⁶). ¹³C NMR spectrum, δ , ppm: 11.2 (Me), 14.2 (CH₂), 25.5 (OCH₂), 71.7 (CH₂C), 119.4, 120.5, 123.7, 136.2, 148.8, 154.0, 156.5. Mass spectrum, m/z ($I_{\rm rel}$, %): 203 (17) M⁺, 119 (28), 105 (52), 79 (100), 78 (70), 52 (20), 51 (37).

Benzaldehyde *O*-[3-(5-tetrazolyl)propyl]oxime (IX). To a suspension of 1.06 g (5.6 mmol) of aldehyde oxime V and 0.14 g (0.56 mmol) of Bu₂SnO in 2 ml of anhydrous toluene was added 1.3 g (11.3 mmol) of azidotrimethylsilane. The reaction mixture was stirred for 30 h at 100°C till disappearance of compound V (GLC monitoring). Then the reaction mixture was neutralized with a saturated solution of NaHCO₃, the precipitate was filtered off and dried in air, and then recrystallized from ethyl acetate. Yield 1 g (43%), colorless crystals, mp 90-92°C. ¹H NMR spectrum, δ , ppm: 2.10 m (2H, CH₂), 2.97 t (2H, OCH₂, J 8.0 Hz), 4.17 t (2H, CH₂CN, J 6.4 Hz), 7.41 m (3H, H³⁻⁵), 7.61 m (2H, H^{2,6}), 8.22 s (1H, CH). 13 C NMR spectrum, δ , ppm: 19.9 (CH₂), 27.0 (OCH₂), 72.5 (<u>C</u>H₂C), 126.8, 128.8, 129.9, 132.0, 148.8, 156.5. Found, %: C 56.81; H 5.62; N 30.01. C-11H₁₃N₅O. Calculated, %: C 57.13; H 5.67; N 30.28.

Acetophenone *O*-[3-(5-tetrazolyl)propyl]oxime (**X**) was prepared in the same way in a 39% yield. On recrystallization from ethyl ether mp 68–70°C. 1 H NMR spectrum, δ, ppm: 2.17 s (3H, Me), 2.24 m (2H, CH₂), 3.18 t (2H, OCH₂, J 7.2 Hz), 4.26 t (2H, CH₂CN,

738 RUBINA et al.

J 5.6 Hz), 7.34 m (3H, H³⁻⁵), 7.56 m (2H, H²⁻⁶). ¹³C NMR spectrum, δ, ppm: 13.0 (Me), 20.3 (CH₂), 27.4 (OCH₂), 72.1 (<u>C</u>H₂C), 126.0, 128.6, 129.4, 136.2, 156.3. Found, %: C 58.68; H 6.15; N 28.51. C₁₂H₁₅N₅O. Calculated, %: C 58.76; H 6.16; N 28.55.

2-Pyridinealdehyde *O*-[3-(5-tetrazolyl)propyl)**oxime** (XI) was prepared in the same way in a 43% yield. The precipitate was recrystallized from a mixture of ethyl acetate and ethanol, mp ~40°C. The compound is unstable and decomposes in air. 1 H NMR spectrum, δ, ppm: 1.99 m (2H, CH₂), 2.71 t (2H, OCH₂, *J* 7.6 Hz), 4.21 t (2H, CH₂CN, *J* 6.6 Hz), 7.42 m (1H, H⁵), 7.84 m (2H, H^{3,4}), 8.16 s (1H, CH), 8.31 s (1H, NH), 8.59 m (1H, H⁶). 13 C NMR spectrum, δ, ppm: 21.7 (CH₂), 28.5 (OCH₂), 74.0 (<u>C</u>H₂C), 120.4, 124.3, 136.9, 148.9, 149.6, 161.7. Found, %: C 51.61; H 5.18; N 36.10. C₁₀H₁₂N₆O. Calculated, %: C 51.72; H 5.21; N 36.19.

2-acetylpyridine *O*-[3-(5-tetrazolyl)propyl]oxime sodium salt (XII) was similarly prepared. The precipitate was recrystallized from a mixture of ethyl acetate and ethanol. Yield 0.86 g (65%), colorless crystals, mp 101–102°C. 1 H NMR spectrum, δ, ppm: 2.01 m (2H, CH₂), 2.25 s (3H, Me), 2.73 t (2H, OCH₂, *J* 7.6 Hz), 4.23 t (2H, CH₂CN, *J* 6.7 Hz), 7.39 m (1H, H⁵), 7.84 m (2H, H^{3,4}), 8.59 m (1H, H⁶). 13 C NMR spectrum, δ, ppm: 10.9 (Me), 21.8 (CH₂), 28.7 (OCH₂), 73.8 (<u>C</u>H₂C), 120.0, 124.0, 136.6, 148.8, 153.5, 154.9, 160.3. Found, %: C 40.81; H 6.18; N 26.10. C₁₁H₂₀N₆NaO₄. Calculated, %: C 40.86; H 6.24; N 25.99.

Single crystals of compound **XII** triclinic, parameters of the unit cell: a 7.8108(3), b 9.6847(4), c 11.6714(6) Å, α 76.096(2), β 83.396(2), γ 77.316(2) deg, V 834.29(6) Å³, F(000) 340, μ 0.12 mm⁻¹, d_{calc} 1.283 g/cm³, Z 2, space group P $\overline{1}$.

Intensities of 6590 reflections (4087 among them independent) were measured on an automatic diffractometer Nonius KappaCCD (molybdenum radiation, λ 0.71073, graphite monochromator) till $2\theta_{max}$ 55°. In calculations were used 2620 reflections with $I > 3\sigma(I)$. The structure was solved by procedure [21]. The refinement was performed by least-squares procedure in the full-matrix anisotropic approximation using software *maxus* [22]. All hydrogen atoms were localized by difference

synthesis and refined in isotropic approximation. The final value of the divergence factor is R 0.043.

REFERENCES

- 1. Koldobskii, G.I., Izv. Akad. Nauk, Ser. Khim., 1995, p. 2115.
- 2. Butler, R.N., *Compr. Heterocycl. Chem. II*, Storr, R.C., Ed., Oxford: Elsevier, 1996, vol. 4, pp. 621, 905.
- 3. Turnbull, K., *Progr. Heterocycl. Chem.*, 1999, 11, p. 163.
- 4. Ostrovskii, V.A. and Koren, A.O., *Heterocycles*, 2000, vol. 53, p. 1421.
- 5. Herr, R.J., Bioorg. Med. Chem., 2002, vol. 10, 3379.
- 6. Sugimoto, Kanaoka, Y., Japan Patent 2001131158; *Chem. Abstr.*, 2001, vol. 134, 353310.
- 7. Chan, Y.P. and Aspnes, G.E., Japan Patent 2001226359; *Chem. Abstr.*, 2001, vol. 135, 180771.
- 8. Aoki, K., Aikawa, K., Kawakami, M., and Yan, Y., WO Patent 0104110; *Chem. Abstr.*, 2001, vol. 134, 115956.
- 9. Forssmann, W.-G., Drexler, H., Walden, M., Schieffer, B., and Schmidt, B., WO Patent 0182858; *Chem. Abstr.*, 2001, vol. 135, 339245.
- 10. Momose, Y., Maekawa, T., Odaka, H., Ikeda, H., and Sohda, T., *Chem. Pharm. Bull.*, 2002, vol. 50, p. 100.
- 11. Druzgala, P., Milner, P. G., and Pfister, J.R., WO Patent 0324943; *Chem. Abstr.*, 2003, vol. 138, 271671.
- 12. Pellicciari, R., Europe Patent 648759; *Chem. Abstr.*, 1995, vol. 123, 256721.
- 13. Connor, D.T. and Mullican, M.D., US Patent 4764525; *Chem. Abstr.*, 1988, vol. 109, 211063.
- 14. Uchida, M., Komatsu, M., Morita, S., Kanbe, T., and Nakaga-wa, K., *Chem. Pharm. Bull.*, 1989, vol. 37, p. 322.
- Kumar, B. and Tilak, D., *Indian J. Chem.*, 1987, vol. 26B, p. 599; *Chem. Abstr.*, 1988, vol. 108, 150379.
- 16. Holbova, E., Sidoova, E., and Uher, M., Czech Patent 217456; *Chem. Abstr.*, 1985, vol. 102, 6499.
- 17. Kraatz, U., Homeyer, B., and Becker, B., Patent 3328273; *Chem. Abstr.*, 1985, vol. 103, 178262.
- 18. Wittenberger, S.J. and Donner, B.G., *J. Org. Chem.*, 1993, vol. 58, p. 4139.
- 19. Kuleshova, L.N. and Zorkii, P.M., *Acta Crystallogr.*, 1981, vol. B37, p. 1363.
- 20. Abele, E., Popelis, Yu., Lukevits, E., Shimanska, M., and Gol'dberg, Yu., *Khim. Geterotsikl. Soedin.*, 1994, p. 18.
- 21. Mishnev, A.F. and Belyakov, S.V., *Kristallografiya*, 1988, vol. 33, p. 835.
- 22. Mackay, S., Gilmore, C.J., Edwards, C., Stewart, N., and Shankland, K., *maXus Computer Program for the Solution and Refinement of Crystal Structures*, Bruker Nonius, Glasgow, 1999.